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Xie et al.

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(54) **NA/K-ATPASE LIGANDS, OUABAIN ANTAGONISTS, ASSAYS AND USES THEREOF**

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(71) Applicant: **The University of Toledo**, Toledo, OH (US)

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(*) Notice: Subject to any disclaimer, the term of this patent is extended or adjusted under 35 U.S.C. 154(b) by 0 days.

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(65) **Prior Publication Data**

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(Continued)

Related U.S. Application Data

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Primary Examiner — Anna Pagonakis

(60) Provisional application No. 61/243,036, filed on Sep. 16, 2009.

(74) *Attorney, Agent, or Firm* — MacMillan, Sobanski & Todd, LLC

(51) **Int. Cl.**

A61K 31/353 (2006.01)

A61K 31/35 (2006.01)

A61K 31/352 (2006.01)

(57) **ABSTRACT**

(52) **U.S. Cl.**

CPC **A61K 31/353** (2013.01); **A61K 31/35** (2013.01); **A61K 31/352** (2013.01)

The present disclosure relates to methods of inhibiting the ATPase activity of Na/K-ATPase without stimulating the receptor function, methods of blocking Na/K-ATPase interaction with Src, methods of inhibiting cell growth, and methods of abolishing ouabain-provoked signaling transduction in the heart of a subject, which includes providing an effective amount of at least one hydroxyl xanthone derivative.

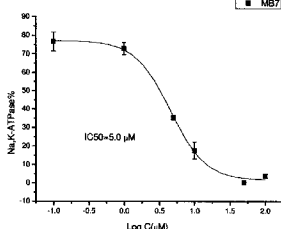
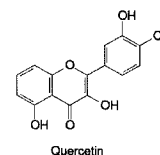
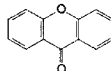
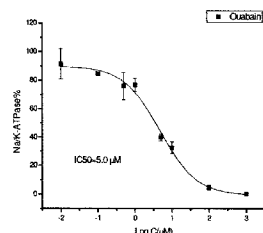
(58) **Field of Classification Search**

None

See application file for complete search history.

4 Claims, 13 Drawing Sheets

(9 of 13 Drawing Sheet(s) Filed in Color)



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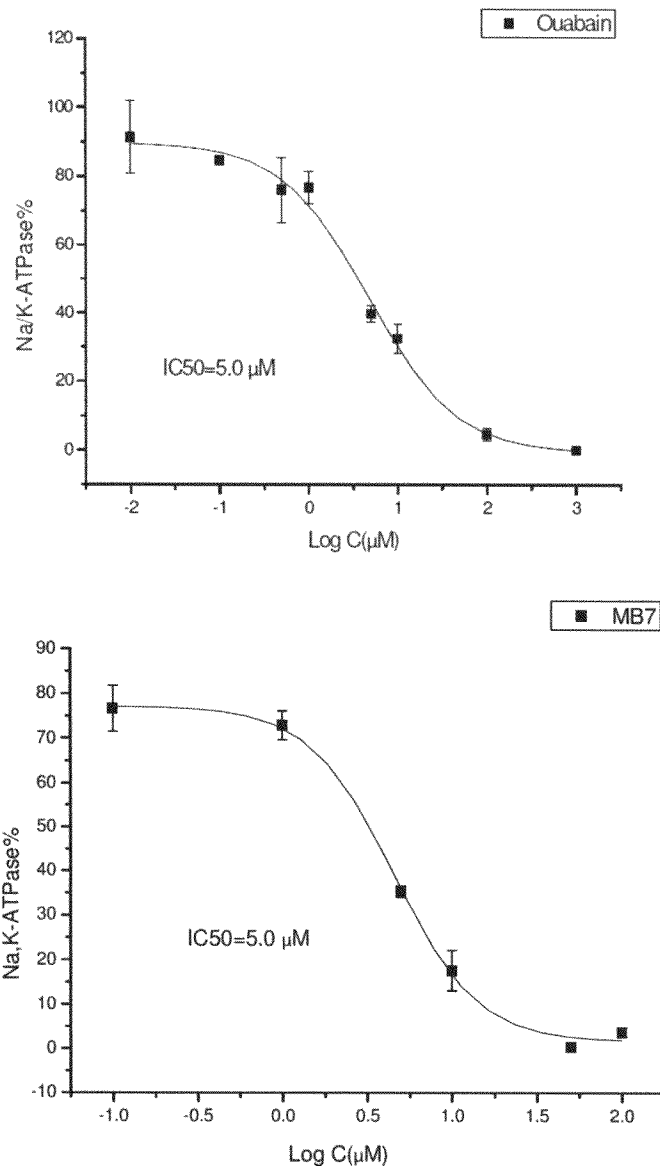


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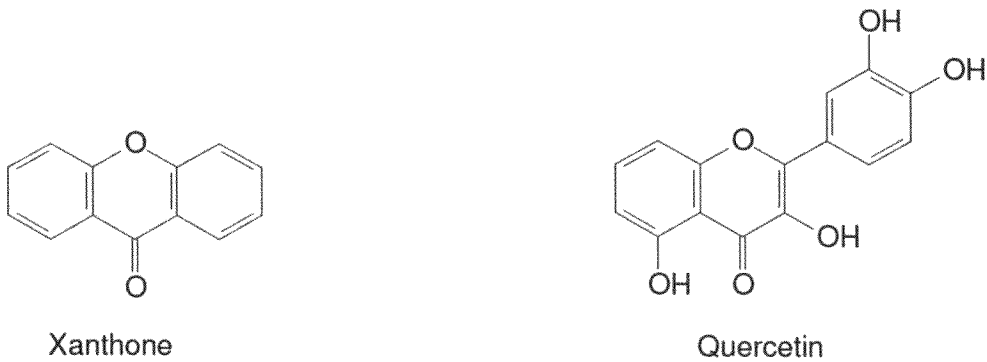


Figure 1B

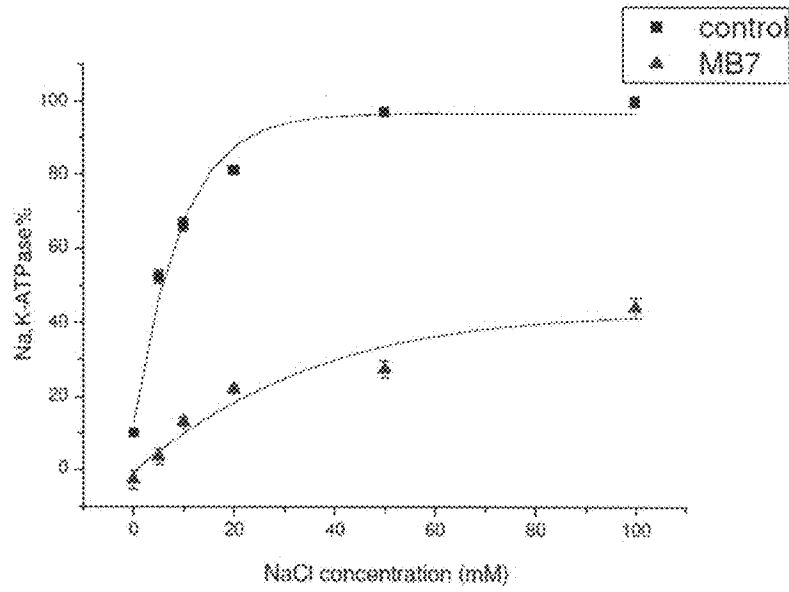


Figure 2A

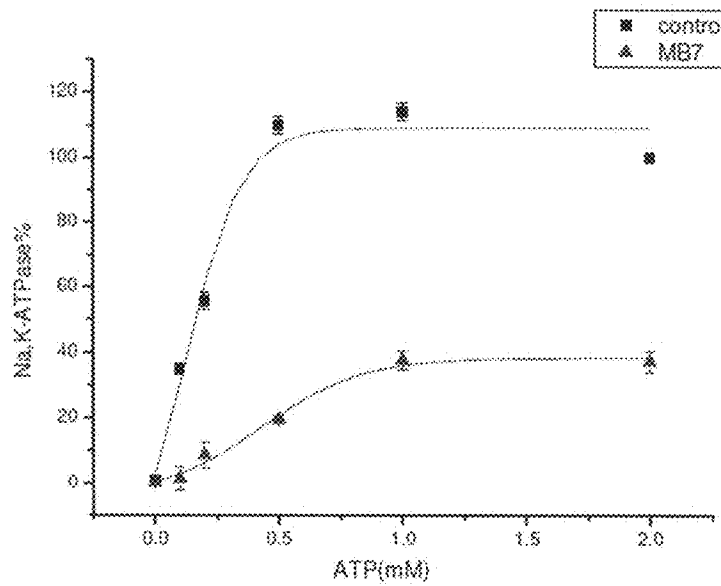


Figure 2B

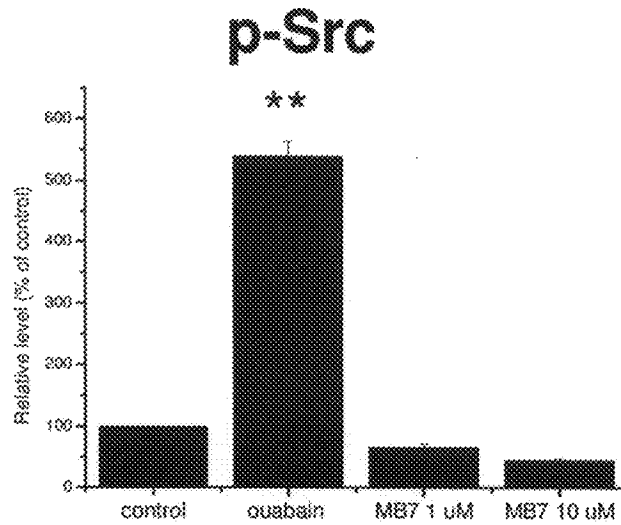
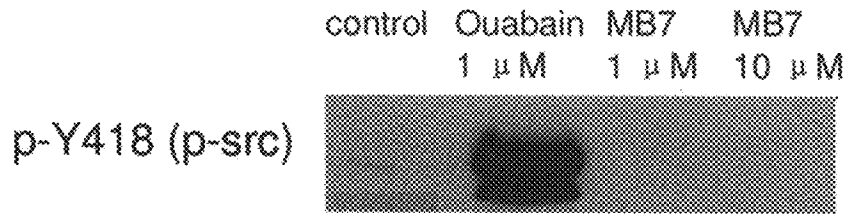


Figure 3

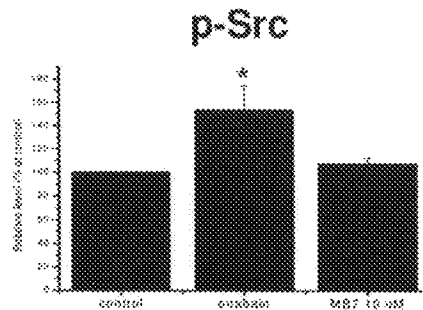
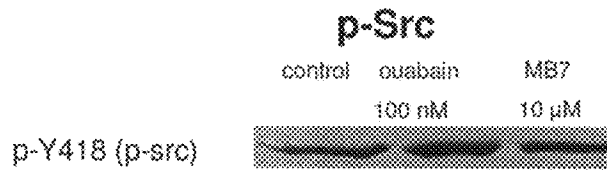


Figure 4A

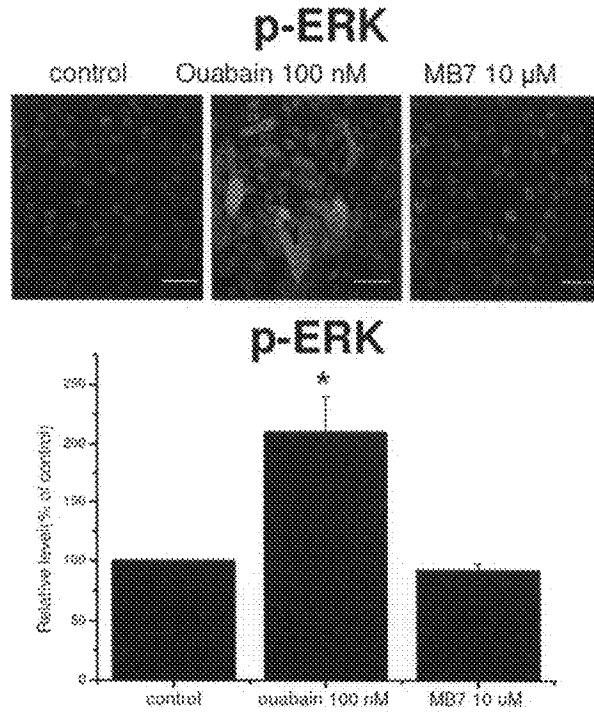


Figure 4B

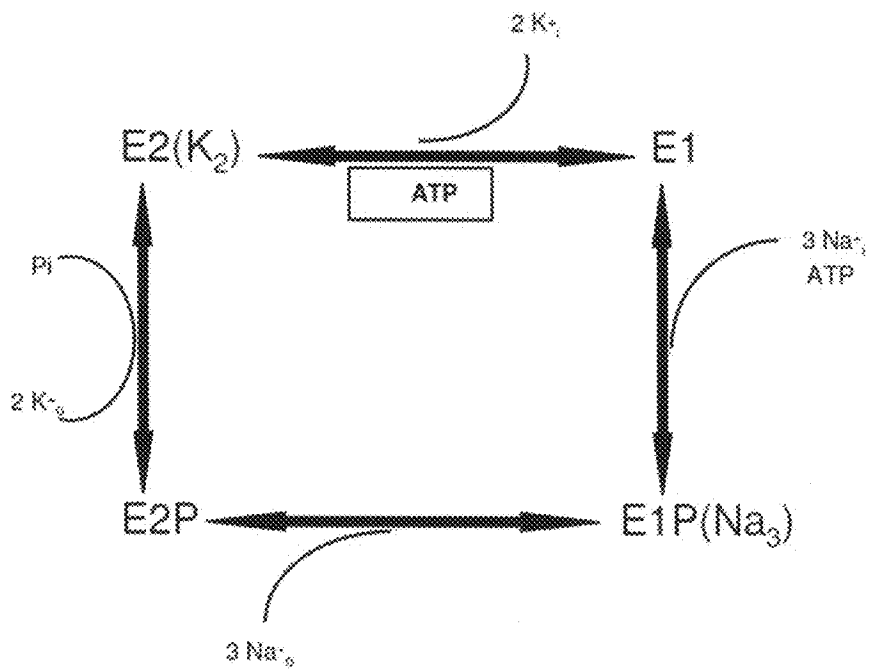


Figure 5A

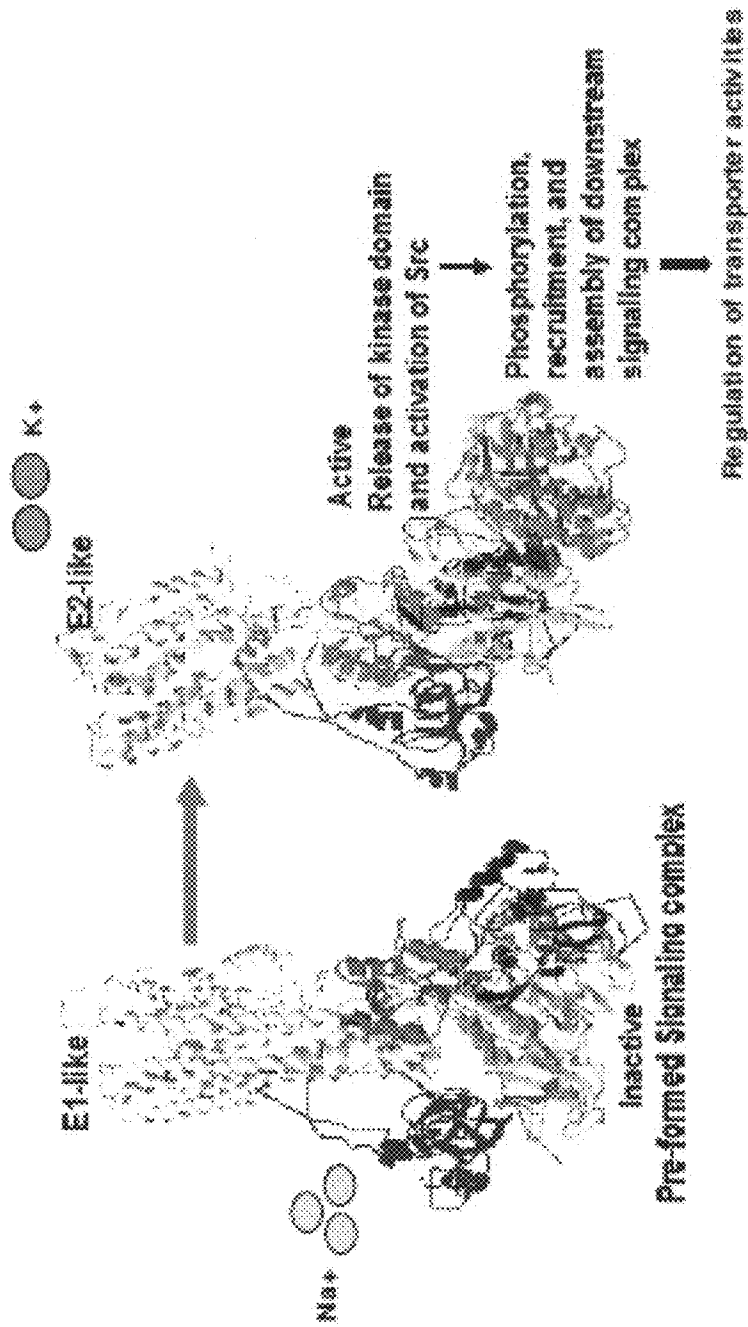


Figure 5B

NaCl (mM)	0	150
KCl(mM)	5	5

pY418

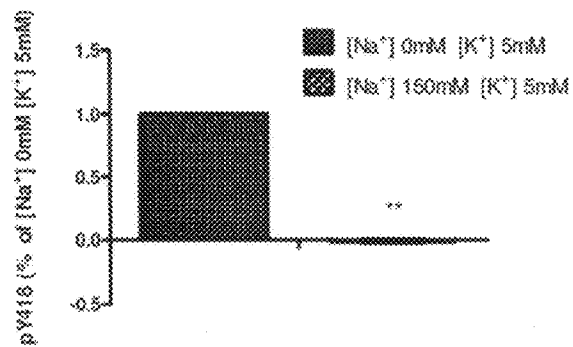


Figure 6A

NaCl (mM)	15	15	15
KCl (mM)	0	1	5

pY418



Figure 6B

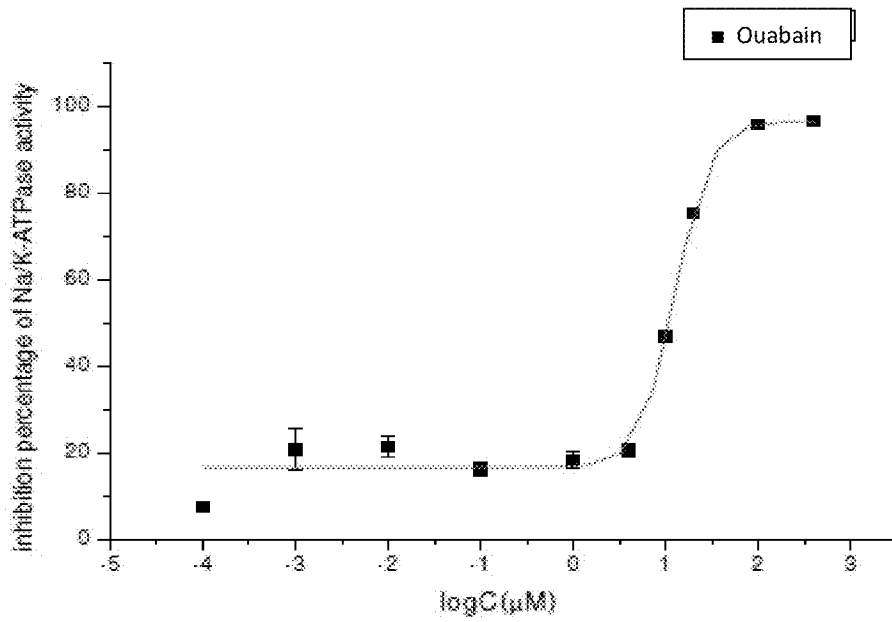


Figure 7A

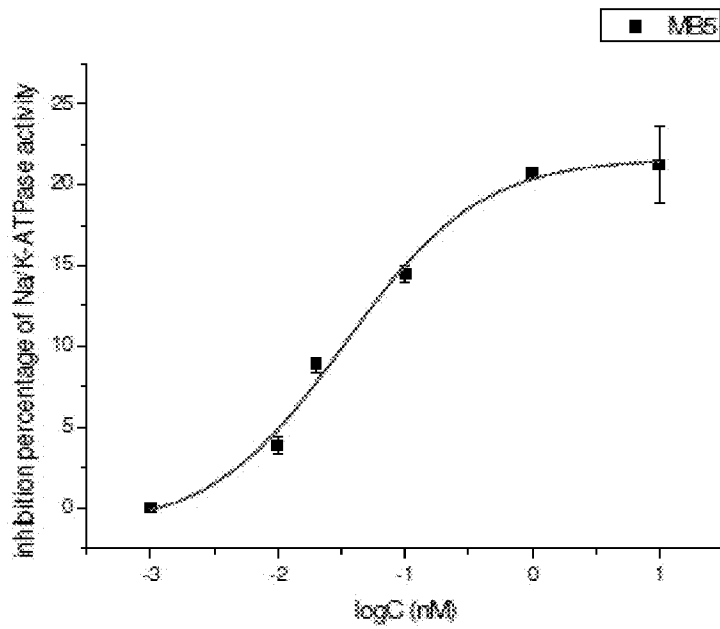


Figure 7B

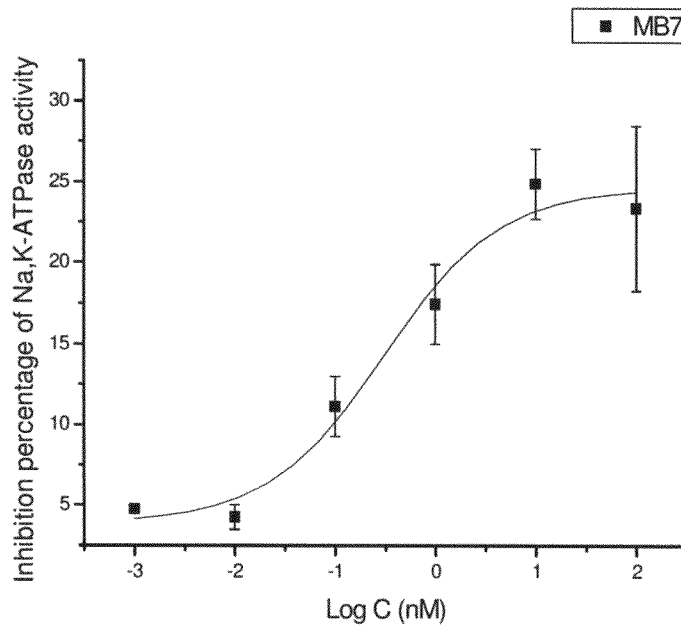


Figure 7C

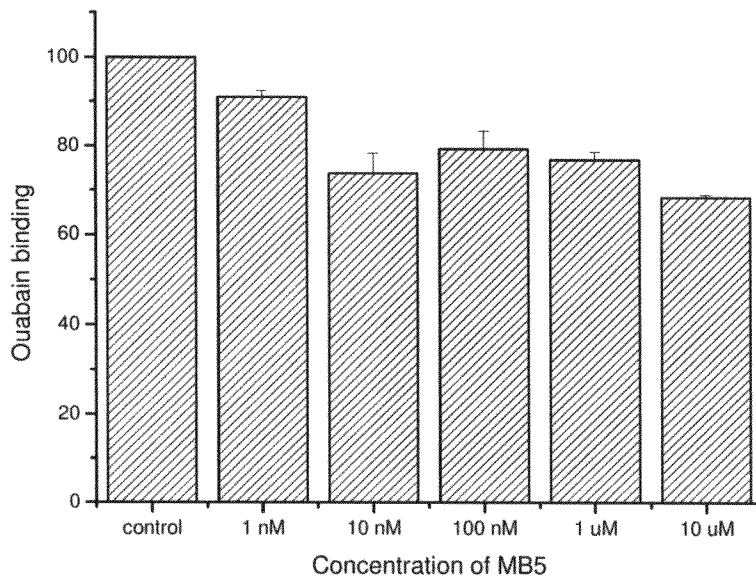
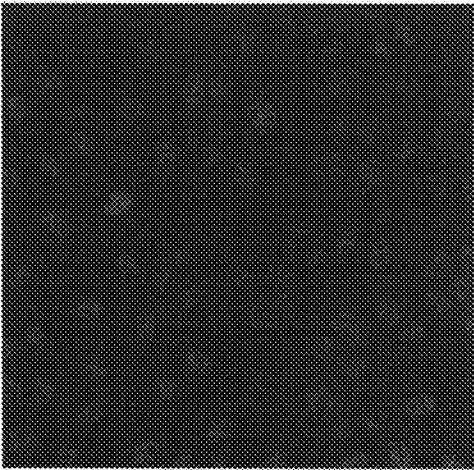
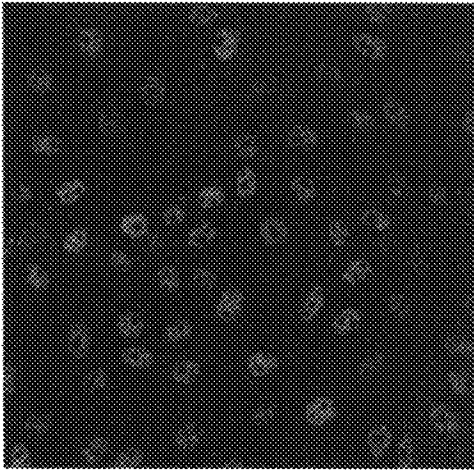


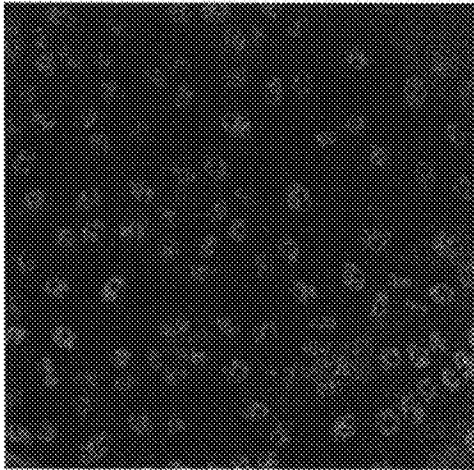
Figure 8



MB5 10 uM 10min



MB5 1nM 10min



control

Figure 9A

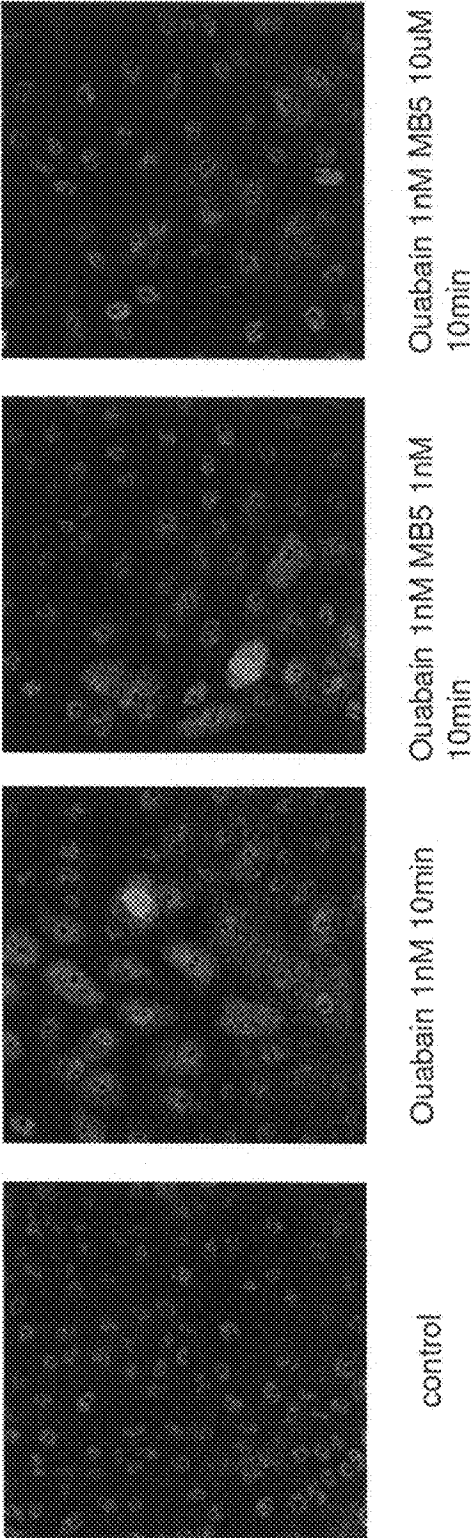


Figure 9B

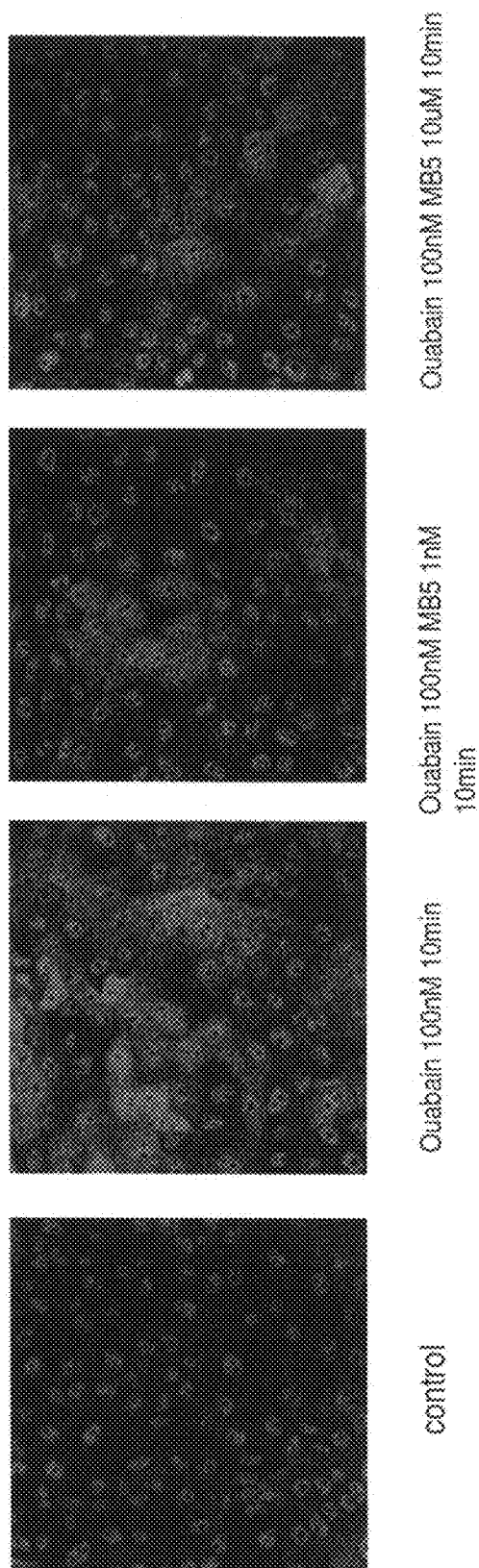


Figure 9C

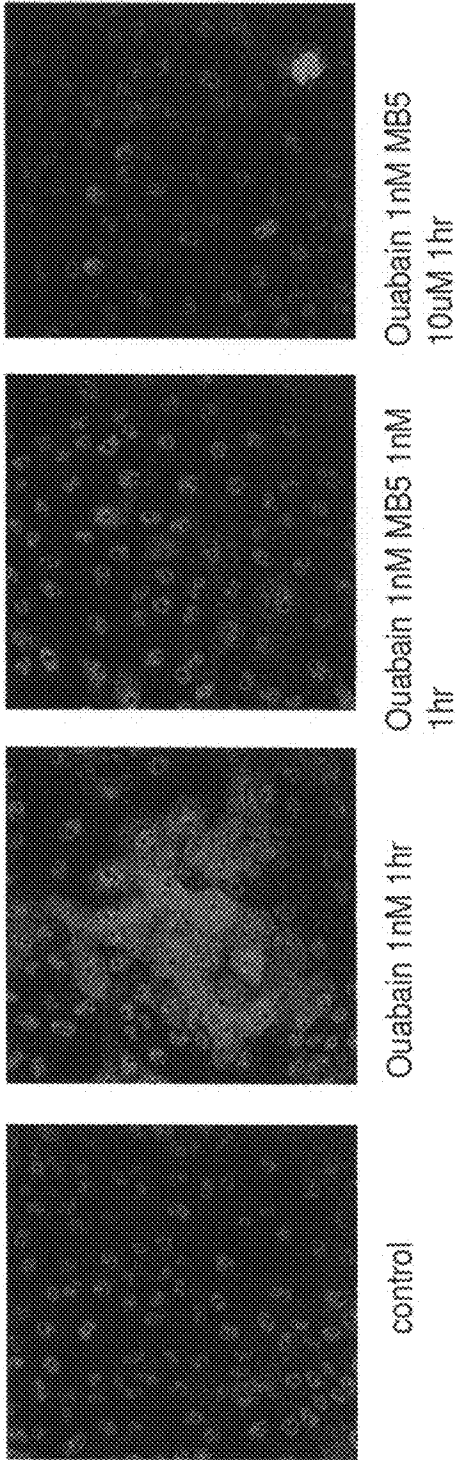


Figure 9D

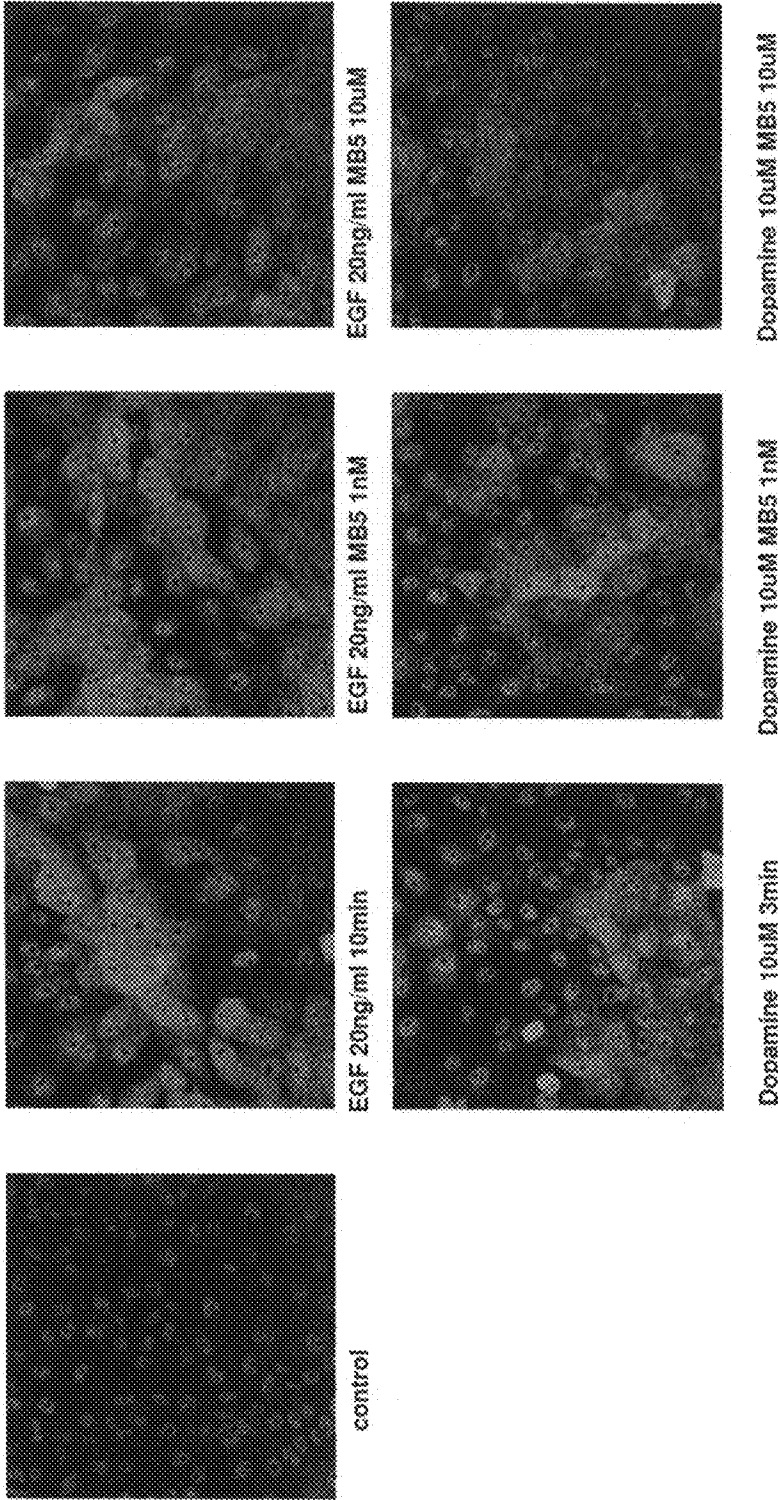


Figure 9E

**NA/K-ATPASE LIGANDS, OUBAIN
ANTAGONISTS, ASSAYS AND USES
THEREOF**

CROSS-REFERENCE TO RELATED
APPLICATIONS

This application is filed under 35 U.S.C §111(a) as a divisional application which claims priority under 35 U.S.C. §119, 35 U.S.C. §120 and the Patent Cooperation Treaty to: parent application U.S. Ser. No. 13/395,700 filed under 35 U.S.C. §371 on Jun. 14, 2012, currently pending; which claims priority to PCT/US2010/048227 filed under the authority of the Patent Cooperation Treaty on Sep. 9, 2010, published; which claims priority to U.S. Provisional Application Ser. No. 61/243,036 filed under 35 U.S.C. §111(b) on Sep. 16, 2009; the disclosures of all priority applications are incorporated herein by reference.

STATEMENT REGARDING FEDERALLY
SPONSORED RESEARCH

This invention was made with government support under GM78565, HL36573 and 2007DFA31370 awarded by the National Institutes of Health. The Government has certain rights in the invention.

TECHNICAL FIELD AND INDUSTRIAL
APPLICABILITY

The present invention directed to Na/K-ATPase ligands and uses thereof. The present invention is also directed to methods and kits for screening substances for agonist and/or antagonist activity of Na/K-ATPase and for treating or preventing diseases or disorders with substances identified by the screening methods described herein.

BACKGROUND

The Na/K-ATPase, also known as the sodium pump, is a ubiquitous trans-membrane enzyme that transports Na⁺ and K⁺ across the plasma membrane by hydrolyzing ATP. It belongs to the family of P-type ATPase that transits between E1 and E2 conformational states during pumping cycles. The functional enzyme is mainly composed of α and β subunits. The α subunit is the catalytic component of the holoenzyme as it contains both the nucleotide and the cation binding sites. Interestingly, studies during the past few years have uncovered many non-pumping functions of Na/K-ATPase such as signal transduction. Specifically, the signaling Na/K-ATPase resides in caveolae and interacts with a number of signaling proteins such as Src, IP3 receptor and caveolin-1. While the interaction between Na/K-ATPase and IP3 receptor facilitates Ca²⁺ signaling, the dynamic association between Na/K-ATPase and Src regulates cellular Src activity and makes it possible for cardiotonic steroids to stimulate protein kinase cascades.

Cardiotonic steroids (CTS) include plant-derived digitalis drugs such as digoxin and ouabain, and vertebrate-derived aglycones such as bufalin and marinobufagenin (MBG).

Although CTS have been considered only as drugs since their discovery, recent studies have identified both ouabain and MBG as endogenous steroids whose production and secretion are regulated by multiple stimuli including angiotensin II and adrenocorticotrophic hormone (ACTH). Of the circulating CTS that have been identified and characterized, ouabain remains the most studied. Moreover, the concentra-

tions of CTS were markedly increased under clinical conditions of high salt loading, chronic renal failure (CRF), and congestive heart failure (CHF). Clinically, digitalis drugs can be used to treat congestive heart failure because they have the well-documented inotropic effects on the heart.

Clinically, these steroids can be used to treat congestive heart failure because they have the well-documented inotropic effects on the heart. It is known that the Na/K-ATPase serves as a receptor for these steroids. While binding of CTS to the Na/K-ATPase inhibits the pumping function, it stimulates the signaling function of Na/K-ATPase. For example, binding of ouabain to the Na/K-ATPase/Src receptor complex stimulates Src kinase. The activated Src, in turn, trans-activates receptor tyrosine kinases such as EGF receptor (EGFR) and converts the tyrosine kinase signal to the stimulation of serine/threonine kinases, lipid kinases and lipases as well as increased production of reactive oxygen species (ROS). Interestingly, while inhibition of Na/K-ATPase by CTS is essential for these drugs to increase cardiac contractile function, stimulation of protein kinases and subsequent increases in the production of ROS by these steroids also cause cardiac hypertrophy and fibrosis in animal studies.

Several of the co-inventors herein have discovered "Na⁺/K⁺-ATPase Ligand," as disclosed in the pending application U.S. Ser. No. 12/087,976 filed Jul. 31, 2008 (claiming priority from PCT/US07/002,365, filed Jan. 30, 2007 (Pub. No. WO 2007/089688 on Aug. 9, 2007), claiming priority from U.S. Ser. No. 60/763,783 filed Jan. 31, 2006), which applications are expressly incorporated herein by reference.

Several of the co-inventors herein have discovered "Na⁺/K⁺-ATPase-Specific Peptide Inhibitors/Activators of Src and Src Family Kinases," as disclosed in the pending application U.S. Ser. No. 12/446,856 filed Apr. 23, 2009 (claiming priority from PCT/US07/023,011, filed Oct. 17, 2007 (Pub. No. WO 2008/054792 on May 8, 2008), claiming priority from U.S. Ser. No. 60/855,482 filed Oct. 16, 2006) which are expressly incorporated herein by reference.

Also, several of the co-inventors herein have discovered "Methods Regulating Na⁺/K⁺-ATPase Expression and Uses thereof. As Therapy for Cancer," as disclosed in the application U.S. Ser. No. 61/109,386 filed Oct. 28, 2008, which is expressly incorporated herein by reference.

Also, several of the co-inventors herein have discovered "Na⁺/K⁺-ATPase-derived Peptides as Antagonists of CTS and as Therapeutic Agents for Cancer," as disclosed in the pending application U.S. Ser. No. 61/122,205, filed Dec. 12, 2008, which is expressly incorporated herein by reference.

SUMMARY

In a broad aspect there is provided herein a novel class of compounds that are different from cardiotonic steroids (CTS), where such compounds inhibit Na/K-ATPase without activating protein kinases.

In another broad aspect, there is provided herein class of compounds comprising Na/K-ATPase ligands which regulate the ion pumping function of Na/K-ATPase.

In another broad aspect, there is provided herein class of compounds comprising Na/K-ATPase ligands which antagonize CTS-induced activation of protein kinase cascades.

In another broad aspect, there is provided herein one or more assays using Na/K-ATPase ligands.

Various objects and advantages of this invention will become apparent to those skilled in the art from the following

detailed description of the preferred embodiment, when read in light of the accompanying drawings.

BRIEF DESCRIPTION OF THE DRAWINGS

The patent or application file may contain one or more drawings executed in color and/or one or more photographs. Copies of this patent or patent application publication with color drawing(s) and/or photograph(s) will be provided by the Patent Office upon request and payment of the necessary fee.

FIG. 1A: Graphs showing concentration curves for ouabain (FIG. 1A-top) and MB7 (3,4,5,6-Tetrahydroxyxanthone) (FIG. 1A-bottom).

FIG. 1B: Chemical structures of xanthone (left) and quercetin (right).

FIGS. 2A-2B: Effects of MB7 on Na⁺ (FIG. 2A) and ATP (FIG. 2B) dependence. Na/K-ATPase activity was measured as described in "Experimental Procedures" as a function of Na⁺ or ATP concentration. MB7 was used at 10 μM.

FIG. 3: Effects of MB7 and ouabain on receptor Na/K-ATPase/Src complex. The purified Na/K-ATPase (2 μg) and purified Src were incubated in the presence of either 10 μM ouabain or 1 and 10 μM MB7 for 15 min and assayed for Src activation as described in "Experimental Procedures". Values are mean SE of at least three independent experiments. **p<0.01 compared with control.

FIG. 4A: Effects of MB7 and ouabain on Src and ERKs. A549 cells were treated with ouabain or MB7 for 10 min and cell lysates (50 μg/lane) were separated by SDS-PAGE and analyzed for Src activation as in FIG. 3. The values are mean±S.E. from four separate experiments. *, p<0.05 versus control.

FIG. 4B: LLC-PK1 cells were treated with MB7 or ouabain for 10 minutes and were immunostained with the ERK/MAPK (Phospho-Thr202/Tyr204) Phosphorylation/Translocation Cell-Based Assay Kit according to the manufacturer's instructions. The images were collected as described under "Experimental Procedures." The scale bar represents 50 μM. Images from a representative experiment are shown. The quantitative data of p-ERK were collected from 40 different fields in three independent experiments and expressed as mean±S.E. *, p<0.05 versus control.

FIG. 5A: Albers-Post Scheme for the Na/K-ATPase pumping cycle.

FIG. 5B: Modeling of the Na/K-ATPase/Src interaction. The A domain (N-terminus and CD2) in Na/K-ATPase was labeled in blue, P domain in green, N domain in black. The SH2 domain of Src was labeled in orange, kinase domain in light blue

FIG. 6A: Effects of changes in Na⁺ and K⁺ concentrations on the receptor Na/K-ATPase/Src activity. Purified Na/K ATPase (2 ug) was resuspended in Tris/HCl (pH7.4) buffer with indicated ion concentrations and incubated with purified c-Src for 15 minutes. Then 3 mM Mg²⁺/ATP was added to the reaction mixture and incubated for another 10 minutes. The samples were analyzed by Western Blot. Values are mean±SE of four separate experiments

FIG. 6B: Effects of K⁺ on the receptor Na/K-ATPase/Src activity. The assay was conducted as in FIG. 6A in the presence of 15 mM NaCl and different concentrations of KCl as indicated. A representative Western blot of three separate experiments is shown.

FIGS. 7A-7C: Dose-dependent inhibition of Na/K-ATPase by MB5: FIG. 7A: The two phase inhibition of both low and high affinity. FIGS. 7B-7C: The dose-response of high affinity inhibition by MB5 (FIG. 7B) and MB7 (FIG. 7C), noting the MB5 is more potent than MB7.

FIG. 8: Effects of MB5 on ouabain binding to the purified Na/K-ATPase. The purified Na/K-ATPase (2 μg) was incubated with 20 nM ³H ouabain in the presence of different concentrations of MB5.

FIG. 9A: Effects of MB5 on ERKs. LLC-PK1 cells were exposed to different concentrations of MB5 (1 nM, 10 μM) for 10 min, and then assayed for active ERKs as described in "Experimental Procedures". A representative set of images is shown. The same experiments were repeated three times.

FIG. 9B: Effects of MB5 on ouabain-induced activation of ERKs. LLC-PK1 cells were pretreated with different concentrations of MB5 (1 nM, 10 μM) for 15 min, exposed to 1 nM ouabain for 10 min, and assayed for active ERKs as in FIG. 9A. A representative set of images is shown.

FIG. 9C: Effects of MB5 on ouabain-induced activation of ERKs. LLC-PK1 cells were pretreated with different concentrations of MB5 (1 nM, 10 μM) for 15 min, exposed to 100 nM ouabain for 10 min, and assayed for active ERKs as in FIG. 9A. A representative set of images is shown.

FIG. 9D: Effects of MB5 on ouabain-induced activation of ERKs. LLC-PK1 cells were pretreated with different concentrations of MB5 (1 nM, 10 μM) for 15 min, exposed to 1 nM ouabain for 1 hour, and assayed for active ERKs as in FIG. 9A. A representative set of images is shown.

FIG. 9E: Effects of MB5 on stimuli-induced activation of ERKs. LLC-PK1 cells were pretreated with different concentrations of MB5 (1 nM, 10 μM) for 15 min, exposed to epidermal growth factor (EGF) or Dopamine stimuli for either 10 or 3 min, and assayed for active ERKs as in FIG. 9A. A representative set of images is shown.

DETAILED DESCRIPTION

Throughout this disclosure, various publications, patents and published patent specifications are referenced by an identifying citation. The disclosures of these publications, patents and published patent specifications are hereby incorporated by reference into the present disclosure to more fully describe the state of the art to which this invention pertains.

The present invention is based at least in part, on the inventors' discovery of a novel molecular mechanism of Na/K-ATPase-mediated Src regulation and the identification of hydroxyl xanthone derivatives that: i) reduce both Na⁺ and ATP affinities on Na/K-ATPase, and ii) act as antagonists that can abolish ouabain-induced activation of kinase cascades.

The present invention is further defined in the following Examples, in which all parts and percentages are by weight and degrees are Celsius, unless otherwise stated. It should be understood that these Examples, while indicating preferred embodiments of the invention, are given by way of illustration only. From the above discussion and these Examples, one skilled in the art can ascertain the essential characteristics of this invention, and without departing from the spirit and scope thereof, can make various changes and modifications of the invention to adapt it to various usages and conditions. All publications, including patents and non-patent literature, referred to in this specification are expressly incorporated by reference. The following examples are intended to illustrate certain preferred embodiments of the invention and should not be interpreted to limit the scope of the invention as defined in the claims, unless so specified.

Example I

Materials

ATP and ouabain were obtained from Sigma (St. Louis, Mo.). Biomol Green was purchased from BIOMOL (Ply-

mouth Meeting, Pa.). ERK/MAPK (Phospho-Thr202/Tyr204) phosphorylation/translocation cell-based assay kit was purchased from Cayman Chemical Company (Ann Arbor, Mich.). Purified recombinant Src was obtained from Upstate Biotechnology (Lake Placid, N.Y.). Polyclonal anti-Tyr(P)418-Src was obtained from Invitrogen (Camarillo, Calif.). Anti-c-Src (B-12) monoclonal antibody was from Santa Cruz Biotechnology Inc. (Santa Cruz, Calif.). The common chemicals were of the highest purity available. Fresh pig kidneys were purchased from a nearby slaughterhouse, and stored at -80°C . until used for enzyme preparation.

High Throughput Screen Assay:

The chemical library used for screening in the present study contained 2600 structurally diverse, drug-like, naturally occurring organic compounds or their semi-synthetic derivatives. Stock compounds were prepared in 96-well plates at 10 mg/ml in DMSO.

The purified Na/K-ATPase was prepared from pig kidney. The specific activities of Na/K-ATPase of various kidney preparations were in the range of 900-1,200 $\mu\text{mol/mg/h}$, which were greater than 95% of the total ATPase activity. The high throughput screen is conducted in a 96-well format with the final reaction volume of 100 μl containing the following components: 100 mM NaCl, 20 mM KCl, 1 mM MgCl_2 , 1 mM EGTA, 20 mM Tris-HCl (pH 7.4) and 0.2 μg of purified Na/K-ATPase. After compounds were added, mixtures were incubated for 15 min at 37°C . for 15 min and reaction was initiated by adding 2 mM ATP.Mg mixture.

Reactions were carried out for 15 min and then stopped by the addition of 100 μl ice-cold trichloroacetic acid. Reaction mixtures were cleared by centrifugation, and assayed for released phosphate using the BIOMOL GREENTM Reagent according to the manufacturer's instructions. In addition, the control Na/K-ATPase activity was measured in the presence and absence of 1 mM ouabain, and taken as 100%. Furthermore, 5 μM ouabain and 0.1% of DMSO were included in each plate as a positive and a vehicle control, respectively. Control experiments showed that ATP hydrolysis catalyzed by the Na/K-ATPase was in linear range within 30 min of incubation under the above experimental conditions.

Cell Culture:

The pig kidney epithelia cells (LLC-PK1 cells) and human lung cancer cells (A549 cells) were obtained from ATCC and maintained in Dulbecco's modified Eagle's medium (DMEM) in the presence of 10% FBS, 100 units/ml penicillin, and 100 $\mu\text{g/ml}$ streptomycin in a 5% CO_2 humidified incubator. To eliminate the confounding effect of growth factors in the serum, cells were serum starved for 24 h before experiments unless otherwise indicated.

Western Blot Analysis:

Cells were washed with PBS and solubilized in ice-cold RIPA buffer containing 1% Nonidet P-40, 1% sodium deoxycholate, 150 mM NaCl, 1 mM EDTA, 1 mM phenylmethylsulfonyl fluoride, 1 mM sodium orthovanadate, 1 mM NaF, 10 $\mu\text{g/ml}$ aprotinin, 10 $\mu\text{g/ml}$ leupeptin, and 50 mM Tris-HCl (pH 7.4) as previously described (13). Cell lysates were then cleared by centrifugation at 14,000 rpm, and supernatants were used for protein assay and subjected to Western blot analysis. Samples were separated on SDS-PAGE (50 $\mu\text{g/lane}$) and transferred to a cellulose membrane. Membranes were blocked with 3% non-fat dried milk for total Src and ERK or 1% BSA plus 1% non-fat dried milk for phosphorylated Src and ERK in TBST (Tris-HCl 10 mM, NaCl 150 mM, Tween 20, 0.1%; pH 8.0) for 1 h at room temperature and then probed with specific antibodies. Protein signals were detected using an ECL kit and quantified using a Bio-Rad GS-670 imaging densitometer.

Assay for the Activation of Receptor Na/K-ATPase/Src Complex:

The activity of receptor Na/K-ATPase/Src complex was assayed. Briefly, the purified Src (4.5 U) was incubated with 2 μg of the purified Na/K-ATPase in phosphate-buffered saline (PBS) for 30 min at 37°C . Afterward, the Na/K-ATPase/Src complex was exposed to ouabain or MB7 for 10 min. Reaction was initiated by addition of 2 mM ATP/Mg²⁺, continued for 5 min at 37°C . and was stopped by addition of SDS sample buffer. The activation of Src was measured by Western blot using anti-pY418 antibody. Total Src was also probed for loading control.

Confocal Imaging and Immunocytochemistry:

LLC-PK1 cells grown on coverslips were serum starved for 24 h and treated with MB7 or ouabain for different times. Immunostaining of p-ERK was performed using the commercially available ERK/MAPK (Phospho-Thr202/Tyr204) Phosphorylation/Translocation Cell-Based Assay Kit according to the manufacturer's instructions. The signals were detected by a Leica confocal microscope. Leica confocal software was used for data analysis.

Data Analysis:

Data are given as mean \pm S.E. Statistical analysis was performed using the Student's t test and significance was accepted at $p < 0.05$.

Results for Example I

High Throughput Screening of Na/K-ATPase Inhibitors

To screen a chemical library for Na/K-ATPase inhibitors, the inventors herein developed a 96-well format assay.

As depicted in FIG. 1A, ouabain, as a positive control, produced a dose-dependent inhibition of Na/K-ATPase. On the other hand, DMSO, the vesicle, showed no effect on the Na/K-ATPase activity when used at concentration below 1% of reaction volume (data not shown). The apparent IC_{50} for ouabain was about 5 μM , comparable to what was reported. The same assay was used to test a total of 2600 compounds at the final concentration of 10 $\mu\text{g/ml}$. This concentration was adapted because a majority of compounds has a molecular mass around 200, thus being tested around 50 μM , ten times of IC_{50} of ouabain. Ouabain (5 μM) was used as a positive control whereas 0.1% DMSO was used as a negative control in each 96-well plate. Assays were conducted in duplicate and the compound that produced at least 25% of inhibition of Na/K-ATPase was identified as a positive hit. Under these experimental conditions, we found a total of 15 positive compounds (Table I below) that include several well-known Na/K-ATPase inhibitors (myricetin, oligomycin, resibufogenin and cinobufagin).

TABLE I

Na/K-ATPase inhibitors identified by high throughput screen		
Sample No.	Name	Formula
2006BD3	Metacycline hydrochloride	$\text{C}_{22}\text{H}_{22}\text{N}_2\text{O}_8 \cdot \text{HCl}$
2006BC4	Mitomycin	$\text{C}_{15}\text{H}_{18}\text{N}_4\text{O}_5$
2008BB8	Tyrothricin	Mixture
2006BC9	4-Epitetracycline	$\text{C}_{22}\text{H}_{24}\text{N}_2\text{O}_8$
2018BF7	Myricetin	$\text{C}_{15}\text{H}_{10}\text{O}_8$
2021BG2	Domiphen bromide	$\text{C}_{22}\text{H}_{40}\text{NO} \cdot \text{Br}$
2035BD1	Resibufogenin	$\text{C}_{24}\text{H}_{32}\text{O}_4$
2035BA5	Cinobufagin	$\text{C}_{26}\text{H}_{34}\text{O}_6$
2036BF1	Oligomycin	$\text{C}_{45}\text{H}_{74}\text{O}_{11}$
MB1	1,3-Dihydroxyxanthone	$\text{C}_{13}\text{H}_8\text{O}_4$

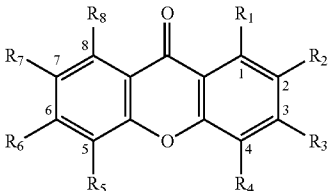
TABLE I-continued

Na/K-ATPase inhibitors identified by high throughput screen		
Sample No.	Name	Formula
MB2	3,4-Dihydroxyxanthone	C ₁₃ H ₈ O ₄
MB3	1,3,5-Trihydroxyxanthone	C ₁₃ H ₈ O ₅
MB5	3,4,5-Trihydroxyxanthone	C ₁₃ H ₈ O ₅
MB6	1,3,5,6-Tetrahydroxyxanthone	C ₁₃ H ₈ O ₆
MB7	3,4,5,6-Tetrahydroxyxanthone	C ₁₃ H ₈ O ₆

Identification of Hydroxyxanthones as a New Class of Na/K-ATPase Ligands:

Among the fifteen positive hits, many are polyphenolic compounds including six hydroxyl xanthone derivatives (MB1 to MB7) (Table 1). Structurally, they are similar to the well-characterized polyphenolic compounds such as quercetin (FIG. 1B). The inventors herein then determined the inhibitory properties of these hydroxyxanthones. Because MB7 was the most potent inhibitor of this group (Table II below), it was used in the following studies.

TABLE II

Structure and Activity Relationship of Xanthone Derivatives									
									
Sample No.	Substituent position								IC ₅₀ (μM)
No.	R ₁	R ₂	R ₃	R ₄	R ₅	R ₆	R ₇	R ₈	(μM)
MB1	OH	H	OH	H	H	H	H	H	>100
MB2	H	H	OH	OH	H	H	H	H	>100
MB3	OH	H	OH	H	OH	H	H	H	65
MB5	H	H	OH	OH	OH	H	H	H	10
MB6	OH	H	OH	H	OH	OH	H	H	60
MB7	H	H	OH	OH	OH	OH	H	H	5
MB8	H	H	OCH ₃	OCH ₃	H	OCH ₃	H	H	>100
2027BA1	OH	H	OCH ₃	H	H	H	OH	OCH ₃	>100
2027BA2	OH	H	OCH ₃	H	H	H	OCH ₃	OH	>100
C-017	OCH ₃	OCH ₃	OCH ₃	H	H	H	OCH ₃	H	>100

In the experiments depicted in FIG. 1A, the dose-response curves of MB7 were compared with ouabain. Like ouabain, MB7 exhibited a dose-dependent inhibition of Na/K-ATPase. The apparent IC₅₀ (5 μM) is comparable to that of ouabain. However, when effects of MB7 on the substrate dependence were determined, MB7, unlike ouabain, reduced both Na⁺ and ATP affinities of Na/K-ATPase (FIG. 2A and FIG. 2B). On the other hand, changes in K⁺ concentration showed no effect on MB7-induced inhibition of Na/K-ATPase (data not shown), but antagonized ouabain-induced inhibition. Furthermore, 10 μM MB7 produced 58±6% inhibition of Na-ATPase as did on Na/K-ATPase. Taken together, the data show that hydroxyxanthones inhibit the Na/K-ATPase through a different mechanism from that of ouabain.

To determine the structure-activity relationship, the dose-response curves of xanthone, six hydroxyxanthones and several methylated hydroxyxanthone derivatives were compared. As depicted in Table II, while xanthone failed to inhibit the Na/K-ATPase activity, an increase in the number of phenolic groups increased the efficacy and potency of hydroxyx-

anthones (e.g., comparing MB2 with MB5, Table II). Consistently, full or partial methylation was able to reduce the inhibitory effect of hydroxyxanthones on the Na/K-ATPase (e.g., comparing MB5 with MB8). Also, when phenolic groups were positioned near the oxygen in the pyrone ring (i.e., at the position 4 and 5), they had more effect on the potency of these compounds (e.g., comparing MB3 with MB5).

MB7 Fails to Activate the Receptor Na/K-ATPase/Src Complex:

The Na/K-ATPase interacts with Src kinase to form a functional receptor complex for ouabain to activate protein kinase cascades.

To determine whether MB7 works as ouabain, capable of activating protein kinases, the inventors herein first measured the effect of MB7 on Src activity using the reconstituted Na/K-ATPase/Src complex. Ouabain (10 μM) was used as a positive control. As depicted in FIG. 3, the Na/K-ATPase inhibited Src. Surprisingly, however, the addition of ouabain, but not MB7, stimulated the Na/K-ATPase-associated Src in

the test tube. This discovery now shows that MB7 can inhibit the ATPase activity without stimulating the receptor function of Na/K-ATPase.

To verify the above results, the effect of MB7 on Src and ERKs in cultured cells was measured. Ouabain was again used as a positive control. As shown in FIG. 4A, while 100 nM ouabain stimulated Src in A549 cells, MB7 up to 10 μM failed to do the same.

To further confirm, LLC-PK1 cells were treated with either 100 nM ouabain or different concentrations of MB7. Ouabain stimulated Src and subsequently the ERK cascade in LLC-PK1 cells. Now, it is shown herein that ouabain increased cellular amount of active ERKs as detected by immunostaining (FIG. 4B). However, under the same experimental conditions, MB7 (from 100 nM to 10 μM) failed to affect cellular ERK activity.

Discussion of Example I

An effective high throughput screen assay was used to identify 15 Na/K-ATPase inhibitors that represent several

structurally divergent classes of compounds. In addition, the newly identified inhibitors were differentiated from other known inhibitors, such as ouabain and oligomycin, by assessing their effects on substrate dependence of the Na/K-ATPase. The newly identified xanthone derivatives, unlike ouabain, only inhibit the ATPase activity, but do not activating the receptor Na/K-ATPase/Src complex.

Xanthone Derivatives as a New Class of Na/K-ATPase Inhibitors:

According to the Albers-Post reaction scheme, the Na/K-ATPase transits from E1 to E2 state via multiple conformational changes. While Na⁺ favors the E1 state, K⁺ promotes the E2 state. Over the years, several classes of organic Na/K-ATPase inhibitors have been identified. They inhibit the Na/K-ATPase by stabilizing the enzyme at different conformational states. For example, CTS bind and stabilize the Na/K-ATPase at the E2P whereas oligomycin prevents E1P to E2P transition.

In comparison, the inventors herein show that the novel ligand, MB7, appears to reduce the formation of E1NaATP because it decreases the apparent affinity of both Na⁺ and ATP without affecting the K⁺ sensitivity of the Na/K-ATPase. Interestingly, xanthenes share the similar chemical structures as flavonoids since they both share the same benzopyrone in their structure (FIG. 1B). Moreover, quercetin and myricetin are well known potent Na/K-ATPase inhibitors. However, unlike MB7, they do not reduce the affinity of Na/K-ATPase towards either Na⁺ or ATP.

The fusion of a benzene ring to benzopyrone completely alters the interaction characteristics of xanthenes with the Na/K-ATPase, which underlines the specificity of xanthone derivative-induced inhibition of Na/K-ATPase.

In addition, the data presented in Table II show the importance of phenolic groups in xanthone derivative-induced inhibition of Na/K-ATPase. The parent compound xanthone has no phenolic group attached to benzene rings and it showed no detectable inhibition of Na/K-ATPase. While tetrahydroxanthone is the most potent inhibitor, dihydroxanthones barely affect the ATPase activity.

While not wishing to be bound by theory, the inventors herein now believe that, since the Na/K-ATPase shares many common features with other P-type ATPases, the newly identified xanthone derivatives described herein may also affect other ion pumps. The inventors herein now also believe that other positive hits such as mitomycin and 4-epitetracycline, are likely to represent another class of new ligands.

The xanthone derivatives compounds have different chemical structures from the known Na/K-ATPase inhibitors, and are now believed by the inventors herein to be a new class of Na/K-ATPase inhibitors that work through a different mechanism.

MB7 does not Activate the Receptor Na/K-ATPase/Src Complex:

The Na/K-ATPase binds Src both in vitro and in vivo. This association regulates cellular Src activity by keeping Src in an inactive state (11). Moreover, the formation of Na/K-ATPase/Src complex produces a functional receptor for ouabain to stimulate the pump-associated Src, which subsequently assembles and activates multiple down stream protein kinase cascades including the ERKs, as shown in FIG. 4.

In contrast to ouabain, the binding of MB7 to the Na/K-ATPase/Src complex in vitro failed to activate Src (FIG. 3). Consistently, it had no effect on cellular Src and ERK activity when MB7 was applied to cultured cells. This discovery shows that MB7, unlike ouabain, fails to alter the conformation of Na/K-ATPase in a way so that the Src kinase domain

can be released from the Na/K-ATPase. The MB7 inhibits Na/K-ATPase in a different way from that of ouabain.

Perspectives:

The name xanthone designates a group of secondary metabolites normally found in a restricted assembly of plants, fungi and lichens. The xanthenes from plants appear to be associated mainly with the families Polygalaceae, Guttiferae, Moraceae and Gentianaceae. These plants have been widely used in traditional Chinese medicine. For example, Yuanzhi, root of *Polygala tenuifolia* Willd. or *Polygala sibirica* L., is extensively used for variety of medical conditions. As phenolic compounds, xanthenes have been described for their antioxidant properties. These properties have been implicated in their anti-inflammatory and chemopreventive actions. One of xanthenes, dimethylxanthenone-4-acetic acid, is currently undergoing clinical trials as an antitumor agent.

As a new class of Na/K-ATPase inhibitors, the xanthenes may also increase cardiac contractile function. Such xanthenes are of great value because they do not stimulate the receptor Na/K-ATPase/Src complex. This becomes important since stimulation of Na/K-ATPase-mediated signal transduction by either endogenous or exogenous CTS alters cardiac growth and induces cardiac fibrosis.

Therefore, MB7 and its analogs are now believed to be especially useful to improve contractile function without precipitating cardiac hypertrophy and/or fibrosis that is often seen under clinical conditions of congestive heart failure.

Example II

Materials

ATP and ouabain were obtained from Sigma (St. Louis, Mo.). Biomol Green was purchased from BIOMOL (Plymouth Meeting, Pa.). ERK/MAPK (Phospho-Thr²⁰²/Tyr²⁰⁴) phosphorylation/translocation cell-based assay kit was purchased from Cayman Chemical Company (Ann Arbor, Mich.). Polyclonal anti-Tyr(P)⁴¹⁸-Src was obtained from Invitrogen (Camarillo, Calif.). Anti-c-Src (B-12) monoclonal antibody was from Santa Cruz Biotechnology Inc. (Santa Cruz, Calif.). The common chemicals were of the highest purity available. Fresh pig kidneys were purchased from a nearby slaughterhouse, and stored at -80° C. until used for enzyme preparation.

Na/K-ATPase Purification and Activity Assay:

The purified Na/K-ATPase was prepared from pig kidney. The specific activities of Na/K-ATPase of various kidney preparations were in the range of 900-1,200 μmol/mg/h, which were greater than 95% of the total ATPase activity. The Na/K-ATPase assay was conducted in the following reaction buffer with the final volume of 500 μl: 100 mM NaCl, 20 mM KCl, 1 mM MgCl₂, 1 mM EGTA, 20 mM Tris-HCl (pH 7.4) and 1 μg of purified Na/K-ATPase. After compounds were added, mixtures were incubated for 15 min at 37° C. for 15 min and reaction was initiated by adding 2 mM ATP.Mg mixture. Reactions were carried out for 15 min and then stopped by the addition of 300 μl ice-cold trichloroacetic acid. Reaction mixtures were cleared by centrifugation, and assayed for released phosphate using the BIOMOL GREEN™ Reagent according to the manufacturer's instructions. Control experiments showed that ATP hydrolysis catalyzed by the Na/K-ATPase was in linear range within 30 min of incubation under the above experimental conditions.

Cell Culture:

The pig kidney epithelia cells (LLC-PK1 cells) and human lung cancer cells (A549 cells) were obtained from ATCC and maintained in Dulbecco's modified Eagle's medium

(DMEM) in the presence of 10% FBS, 100 units/ml penicillin, and 100 µg/ml streptomycin in a 5% CO₂ humidified incubator. To eliminate the confounding effect of growth factors in the serum, cells were serum starved for 24 h before experiments unless otherwise indicated.

Src Activity Assay and Western Blot Analysis:

The protocol to measure the effects of Na/K-ATPase and its ligands on Src activity by Western blotting Src pY 418 phosphorylation were followed. To prepare cell lysates for Western analysis, cells were washed with PBS and solubilized in ice-cold RIPA buffer containing 1% Nonidet P-40, 1% sodium deoxycholate, 150 mM NaCl, 1 mM EDTA, 1 mM phenylmethylsulfonyl fluoride, 1 mM sodium orthovanadate, 1 mM NaF, 10 µg/ml aprotinin, 10 µg/ml leupeptin, and 50 mM Tris-HCl (pH 7.4). Cell lysates were then cleared by centrifugation at 14,000 rpm, and supernatants were used for protein assay and subjected to Western blot analysis. Samples were separated on SDS-PAGE (50 µg/lane) and transferred to a cellulose membrane. Membranes were blocked with 3% non-fat dried milk for total Src and ERK or 1% BSA plus 1% non-fat dried milk for phosphorylated Src and ERK in TBST (Tris-HCl 10 mM, NaCl 150 mM, Tween 20, 0.1%; pH 8.0) for 1 h at room temperature and then probed with specific antibodies. Protein signals were detected using an ECL kit and quantified using a Bio-Rad GS-670 imaging densitometer.

Confocal Imaging and Immunocytochemistry:

LLC-PK1 cells grown on coverslips were serum starved for 24 h and treated with MB5 or ouabain for different times. Immunostaining of p-ERK was performed using the commercial available ERK/MAPK (Phospho-Thr202/Tyr204) Phosphorylation/Translocation Cell-Based Assay Kit according to the manufacturer's instructions. The signals were detected by a Leica confocal microscope. Leica confocal software was used for data analysis.

Data Analysis:

Data are given as mean±S.E. Statistical analysis was performed using the Student's t test and significance was accepted at p<0.05.

Results for Example II

The Na/K-ATPase Regulates Src in a Conformation-Dependent Manner

A large number of Na/K-ATPase interacts directly with Src kinase in life cells. The interaction involves at least two pairs of protein domains. Specifically, the second cytosolic domain of µl subunit interacts with the Src SH2 and the nucleotide binding (N) domain associates with the Src kinase domain. During the pumping cycle, the Na/K-ATPase undergoes E1 to E2 conformational transition (FIG. 5A) where the N domain closes up and the A domain rotates to dock onto the N domain (FIG. 5B).

Structure modeling shows that the location of and the space between the A and N domains only at the E1 state are suitable for the µl subunit to bind both the SH2 and the kinase domains simultaneously (FIG. 5B). Thus, it is now believed by the inventors herein that, while the E1 Na/K-ATPase inhibits Src, the Na/K-ATPase-associated Src must be active when the enzyme is at the E2 conformation.

Indeed, when the majority of Na/K-ATPase is stabilized in the E2 conformation by incubating the enzyme in Na⁺-free and 5 mM buffer (47), the Na/K-ATPase-associated Src became fully active (FIG. 6A). Consistently, addition of 150 mM Na⁺ to this reaction buffer destabilized the E2 Na/K-ATPase, which resulted in the inactivation of Src (FIG. 6A).

Moreover, reduction of K⁺ from 5 mM to 0 caused a gradual activation of Src (FIG. 6B). These results show that the interaction between the Na/K-ATPase and Src can be regulated by changes in the conformational state of Na/K-ATPase. Furthermore, inhibition of E1 to E2 conformational transition of Na/K-ATPase can be used to antagonize ouabain-induced Src activation.

Xanthone Derivatives are Potent Na/K-ATPase Inhibitors and Prevents Ouabain from Binding to the Na/K-ATPase:

New xanthone derivatives as a class of new Na/K-ATPase ligands that stabilize the Na/K-ATPase in the E1 conformation are described in EXAMPLE I.

As depicted in FIGS. 1 and 7A-C, hydroxyxanthones exhibited two phases of inhibition of Na/K-ATPase activity. The high affinity inhibition by MB5 occurs at sub-nM concentrations (IC₅₀ of 40 pM), producing about 25% inhibition of the ATPase activity, whereas low affinity binding produces 100% inhibition at about 50 µM. It is of interest to note that while MB5 is more potent than MB7 at the high affinity site, it has lower potency at low affinity site. Because ouabain prefers to bind the E2 Na/K-ATPase, stabilization of the Na/K-ATPase at the E1 state by MB5 can be used to antagonize ouabain binding. Indeed, as depicted in FIG. 8, MB5 produced a dose-dependent inhibition of ouabain binding to the purified Na/K-ATPase. The significant inhibition was detected at 0.1 nM and maximal inhibition of 30% was reached when the Na/K-ATPase was exposed to 10 µM MB5.

MB5 Antagonizes Ouabain-, but not EGF and Dopamine, Induced Activation of Protein Kinases:

Ouabain stimulates protein kinase cascades via a Src-dependent pathway in LLC-PK1 cells. To determine whether MB5 can antagonize ouabain-induced signal transduction, treated LLC-PK1 cells were pre-treated with different concentrations of MB5 for 15 min and then exposed to ouabain or other stimuli.

As shown in FIGS. 9A-9D, while ouabain (1 nM and 100 nM) stimulated ERKs in a time and concentration-dependent manner, MB5 at 1 nM was able to significantly reduce ouabain-induced activation of ERKs. Moreover, when MB5 was used at 10 µM, it completely blocked the effect of ouabain on ERKs in LLC-PK1 cells.

In contrast, when the same experiments were conducted in the presence of either EGF or dopamine, MB5 failed to affect ERK activation induced by these two stimuli (FIG. 9E).

Taken together, these data show that MB5 can function as a specific ouabain antagonist to against the activation of protein kinase cascades.

Discussion of Example II

The receptor Na/K-ATPase/Src complex can be activated when the Na/K-ATPase is stabilized in the E2 conformation, whereas the E1 Na/K-ATPase keeps Src in an inactive state.

Described herein is an effective high throughput screen assay for chemicals that can regulate both pumping and signaling functions of Na/K-ATPase.

The high throughput assay (as described in EXAMPLE I) is useful to identify chemicals/compositions that affect the E1/E2 conformational transition, as reflected by the inhibition of Na/K-ATPase. The identified chemicals/compositions can then be tested as either agonists or antagonists of Na/K-ATPase/Src receptor complex as follows: As agonists, such chemicals/compositions would function as ouabain and their inhibitory effects on the Na/K-ATPase could be antagonized by increasing K⁺ concentration. On the other hand, as antagonists they would act as MB5, preventing ouabain binding to

13

the Na/K-ATPase. Moreover, their inhibitory effect on Na/K-ATPase could be reduced by increasing Na⁺ concentration.

Also described herein is the identification of a class of novel and potent ouabain antagonists that prevent ouabain from activating the Na/K-ATPase/Src receptor complex. Subsequently, the ouabain antagonists also abolished ouabain-induced activation of ERKs.

These results are significant for at least the following considerations: 1) Na/K-ATPase complexes and/or Na/K-ATPase/Src complexes are useful for screening new agonists and antagonists; 2) Xanthone derivatives, as described herein, are useful as medications to antagonize endogenous CTS elevation-induced pathological changes that include, for example, cardiovascular and renal remodeling as well as cardiac and renal fibrosis; 3) Xanthone derivatives, as described herein, are useful as medications to treat congestive heart failure because they can not only improve contractile function but also prevent structural remodeling in the myocardium; 4) Xanthone derivatives, as described herein, are useful as medications to treat prostate and other cancers because the Na/K-ATPase signaling is important for the growth of these cancer cells; and, 5) Xanthone derivatives are useful for the new generation of receptor Na/K-ATPase antagonists that will be more potent, less toxic and have better pharmacokinetic properties than currently available compositions.

While the invention has been described with reference to various and preferred embodiments, it should be understood by those skilled in the art that various changes may be made

14

and equivalents may be substituted for elements thereof without departing from the essential scope of the invention. In addition, many modifications may be made to adapt a particular situation or material to the teachings of the invention without departing from the essential scope thereof. Therefore, it is intended that the invention not be limited to the particular embodiment disclosed herein contemplated for carrying out this invention, but that the invention will include all embodiments falling within the scope of the claims.

What is claimed is:

1. A method for inhibiting cancer cell proliferation comprising:
 - administering to a subject in need thereof an effective amount of at least one Na/K-ATPase ligand selected from the group consisting of: MB5 (3,4,5-trihydroxyxanthone) and MB7 (3,4,5,6-tetrahydroxyxanthone);
 - and
 - inhibiting the ATPase activity without stimulating the receptor function of Na/K-ATPase in the subject in need of inhibition of cancer cell proliferation.
2. The method of claim 1, wherein the Na/K-ATPase ligand is MB5 (3,4,5-trihydroxyxanthone).
3. The method of claim 1, wherein the Na/K-ATPase ligand is MB7 (3,4,5,6-tetrahydroxyxanthone).
4. The method of claim 2, wherein the Na/K-ATPase ligand is present at 1.0 nM.

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