

[54] **NOVEL ANTAGONISTS OF THE ANTIDIURETIC ACTION OF ARGININE VASOPRESSIN**

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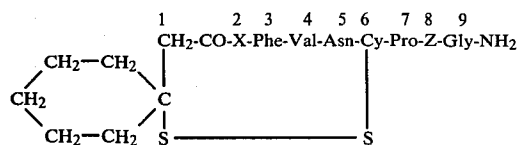
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[57]

ABSTRACT

Compounds acting as antagonists of the antidiuretic activity of arginine vasopressin are those of the formula



wherein X is D-Phe, D-Val, D-Leu, D-Ile, D-Arg, D-norvaline, D-norleucine, D-cyclohexylalanine, D- α -aminobutyric acid, D-threonine or D-methionine and Z is D- or L-Arg.

13 Claims, No Drawings

deamino[2-(O-methyl)-tyrosine]-arginine vasopressin; dPTyr(Me)AVP, [1-deaminopenicillamine, 2-(O-methyl)tyrosine]-arginine vasopressin; d(CH₂)₅Tyr(Me)VDAVP, [1-(β-mercapto-β,β-cyclopentamethylenepropionic acid), 2-O-methyltyrosine, 4-valine, 8-D-arginine] vasopressin; d(CH₂)₅ D-Tyr VDAVP, [1-(β-mercapto-β,β-cyclopentamethylenepropionic acid), 2-D-tyrosine, 4-valine, 8-D-arginine] vasopressin; d(CH₂)₅ D-Tyr VAVP, [1-(β-mercapto-β,β-cyclopentamethylenepropionic acid), 2-D-tyrosine, 4-valine]-arginine vasopressin; d(CH₂)₅D-Phe² VDAVP, [1-(β-mercapto-β,β-cyclopentamethylenepropionic acid), 2-D-phenylalanine, 4-valine, 8-D-arginine] vasopressin; d(CH₂)₅ D-Phe VAVP, [1-(β-mercapto-β,β-cyclopentamethylenepropionic acid), 2-D-phenylalanine, 4-valine]-arginine vasopressin; d(CH₂)₅ [Gly²] VAVP, [1-(β-mercapto-β,β-cyclopentamethylenepropionic acid), 2-glycine, 4-valine]-arginine vasopressin; d(CH₂)₅[D-Ala²] VAVP, [1-(β-mercapto-β,β-cyclopentamethylenepropionic acid), 2-D-alanine, 4-valine]-arginine vasopressin; d(CH₂)₅ [D-Val²] VAVP [1-(β-mercapto-β,β-cyclopentamethylenepropionic acid), 2-D-valine, 4-valine]-arginine vasopressin; d(CH₂)₅ [D-Leu²] VAVP, [1-(β-mercapto-β,β-cyclopentamethylenepropionic acid), 2-D-leucine, 4-valine]-arginine vasopressin; d(CH₂)₅ [D-Ile²] VAVP, [1-(β-mercapto-β,β-cyclopentamethylenepropionic acid), 2-D-isoleucine, 4-valine]-arginine vasopressin; and d(CH₂)₅ [D-Arg²] VAVP, [1-(β-mercapto-β,β-cyclopentamethylenepropionic acid), 2-D-arginine, 4-valine]-arginine vasopressin.

The active peptides were synthesized by solid phase synthesis as described by Bankowski et al. (1978), supra; Merrifield, *J. Am. Chem. Soc.*, vol. 85 (1963) at 2149 and *Biochemistry*, vol. 3 (1964) at 1385; Manning, *J. Am. Chem. Soc.*, vol. 90 (1968) at 1348; Manning et al., *J. Med. Chem.*, vol. 19 (1976) at 376; Lowbridge et al., *J. Med. Chem.*, vol. 20 (1977) at 1173; Manning et al., *J. Med. Chem.*, vol. 16 (1973) at 975; Kruszynski et al. (1980), supra; Sawyer et al., (1981), supra; or Manning et al. (1981), supra.

Initial attempts to design an antagonist of the antidiuretic response to arginine vasopressin (AVP) including synthesis of [1-deaminopenicillamine, 4-valine, 8-D-arginine] vasopressin (dPVDAVP) by Manning et al. (1977), supra, and of [1-(β-mercapto-β,β-cyclopentamethylenepropionic acid), 4-valine, 8-D-arginine] vasopressin (d(CH₂)₅VDAVP), Lowbridge (1978), supra. These analogs were designed by replacing the two hydrogens on the β-carbon at the 1-position of the highly active and selective antidiuretic peptide 1-deamino[4-valine, 8-D-arginine] vasopressin (dVDAVP), Manning et al., *J. Med. Chem.*, vol. 16 (1973) at 975, by two methyl groups and a cyclopentamethylene group, respectively. These substituents had previously been shown to convert the highly potent oxytocic agonist 1-deamino-oxytocin (dOT) into potent antagonists of the oxytocic response to oxytocin, specifically, [1-deaminopenicillamine] oxytocin (dPOT) and [1-(β-mercapto-β,β-cyclopentamethylenepropionic acid)] oxytocin (d(CH₂)₅OT). See, Hope et al., *J. Biol. Chem.*, vol. 237 (1962) at 1563, Schulz et al., *J. Med. Chem.*, vol. 9 (1966) at 647 and Nestor et al., *J. Med. Chem.*, vol. 18 (1975) at 284.

The discovery of the antidiuretic antagonists d(CH₂)₅Tyr(alk)VAVP, Sawyer, et al., (1981), supra, Manning et al., (1981) supra, led to the synthesis of other position two substituted analogs. Enhanced anti-

antidiuretic potencies were exhibited by the various O-alkyl D-tyrosine analogs, Manning et al., in *Peptides, Structure, Function*, Dan H. Rich and E. Gross, eds., Pierce Chemical Co (in press) and *J. Med. Chem.* (in press). The unalkylated D-tyrosine isomers of d(CH₂)₅VDAVP and d(CH₂)₅VAVP, i.e., d(CH₂)₅D-Tyr-VDAVP and d(CH₂)₅D-Tyr-VAVP were also shown to be anti-antidiuretics. Attempts to further enhance anti-antidiuretic potency and selectivity have led to the synthesis of analogs of d(CH₂)₅D-Tyr²VAVP and d(CH₂)₅D-Tyr²VDAVP containing other D-amino acids in place of D-tyrosine at position two, in accordance with the present invention.

It was found, in accordance with the present invention, that some d(CH₂)₅VAVP derivatives having a D-amino acid other than tyrosine and larger than alanine in the 2-position are more potent antagonists of the antidiuretic action of AVP than compounds having D- or L-tyrosine ether units or a D-tyrosine unit at the 2-position of d(CH₂)₅VAVP or d(CH₂)₅VDAVP.

Preferred compounds of this invention are those wherein the 8-substituent is Arg and the 2-substituent is D-Phe, D-Val, D-Leu and D-Ile.

As shown by intravenous administration of the compounds of the invention to hydrated rats anesthetized with ethanol, compounds having D-Phe, D-Val, D-Leu or D-Ile substituents at the 2-position have high pA₂ values and effective doses near or lower than the lowest effective doses known heretofore.

Compounds having D-Phe, D-Val, D-Leu or D-Ile at the 2-position and Arg at the 8-position are also pure antidiuretic antagonists, i.e., these compounds have no transient antidiuretic agonism. Moreover, these compounds are more selective in their activity, by virtue of high anti-ADH/antivasopressor activity ratios, than known compounds.

The compounds of this invention are very effective antagonists of the antidiuretic response to ADH. They can therefore be used in pharmacological studies on the contribution of ADH to a variety of pathological states involving water retention. It is further contemplated that they could be effective and specific agents for treating the syndrome of inappropriate secretion of ADH, that is, the Schwartz-Bartter syndrome or SIADH. This syndrome can complicate a number of disorders, including carcinomas, pulmonary diseases, intracranial diseases and head injuries, Bartter et al., *Am. J. Med.*, vol. 42 (1967) at 790.

The compounds of this invention can be employed in mixture with conventional excipients, i.e., physiologically and pharmaceutically acceptable organic or inorganic carriers suitable for parenteral or enteral application, which do not interact deleteriously with the active compounds.

Suitable pharmaceutically acceptable carriers include, but are not limited to, water, salt solutions, alcohols, vegetable oils, polyethylene glycols, gelatine, lactose, amylose, magnesium stearate, talc, silicic acid, viscous paraffin, perfume oil, fatty acid monoglycerides and diglycerides, pentaerythritol fatty acid esters, hydroxy-methylcellulose, polyvinyl pyrrolidone, etc. The pharmaceutical preparations can be sterilized and if desired mixed with auxiliary agents, e.g., lubricants, preservatives, stabilizers, wetting agents, emulsifiers, salts for influencing osmotic pressure, buffers, coloring, flavoring and/or aromatic substances and the like which do not deleteriously react with the active compounds.

For parenteral or intranasal application, solutions, preferably aqueous solutions, as well as suspensions, emulsions, or implants, including suppositories, are particularly suitable. Ampoules are convenient unit dosages.

The compounds of this invention are generally administered to animals, including but not limited to mammals, e.g., livestock, household pets, humans, cattle, cats and dogs. A diuretically effective daily dosage of the active compounds can be administered parenterally in a single dosage or as divided dosages throughout the day.

Parenteral or intranasal administration is preferred, the compounds of Formula I of this invention being particularly valuable in the treatment of humans afflicted with water retention of any etiology. In this regard, they can be administered in substantially the same manner as the known compounds oxytocin and vasopressin, to achieve their physiological effects.

It will be appreciated that the actual preferred amounts of active compounds used will vary according to the specific compound being utilized, the particular compositions formulated, the mode of application, and the particular organism being treated. Optimal application rates under/in a given set of conditions can be ascertained by those skilled in the art of using conventional dosage determination tests in view of the above guidelines.

DESCRIPTION OF PREFERRED EMBODIMENT

Preferred antidiuretic antagonists of the invention are compounds of Formula I, wherein X is D-Phe, D-Val, D-Leu or D-Ile and Z is L-Arg. The D-Ile or D-Phe compound is most preferred.

Without further elaboration, it is believed that one skilled in the art can, using the preceding description, utilize the present invention to its fullest extent. The following specific embodiments are, therefore, to be construed as merely illustrative and not limitative of the remainder of the disclosure in any way whatsoever. In the following Examples, the temperatures are set forth uncorrected in degrees Celsius. Unless otherwise indicated, all parts and percentages are by weight.

Chloromethylated resin (Bio-Rad Bio-Beads SX-1) was esterified by the procedure of Gisin, *Helv. Chim. Acta.*, vol. 56 (1973) at 1476 with Boc-Gly until 0.47 mmol/g and ~0.64 mmol/g were incorporated. Amino acid derivatives including Boc-Tyr(Me) ($R_f(A)$ 0.7; $R_f(B)$ 0.8) were supplied by Bachem Inc., or synthesized.

Triethylamine (TEA) and N-methylmorpholine (NMM) were distilled from ninhydrin.

Acetic acid used as the HCl-acetic acid cleavage reagent was heated under reflux with boron triacetate and distilled from the reagent. Dimethylformamide (DMF) was distilled under reduced pressure immediately before use. Methanol was dried with magnesium methoxide and distilled. Other solvents and reagents were analytical grade.

Thin layer chromatography (TLC) done on silica gel plates (0.25 mm, Brinkmann Silplate) using the following solvent systems: A. cyclohexane-chloroform-acetic acid (2:8:1 v/v); B. propan-1-ol-ammonia (34%) (2:1 v/v); C. ethanol (95%)-ammonia (34%) (3:1 v/v); D. chloroform-methanol 7:3 v/v; E. butan-1-ol-acetic acid-water (4:1:5 v/v, upper phase); F. butan-1-ol-acetic acid-water-pyridine (15:3:3:10 v/v). The applied loadings were 10–50 μ g. The minimum length of the chromatograms was 10 cm. Chloroplatinate reagent and

iodine vapor were used for development of the chromatograms.

Amino acid analysis of the peptides was done by the method of Spackman et al., *Anal. Chem.* vol. 30 (1958) at 1190, in which peptide samples weighing about 0.5 mg were hydrolyzed with constant boiling hydrochloric acid (400 μ l) in evacuated and sealed ampoules for 18 h at 120° C. The analyses were performed using a Beckman Automatic Amino Acid Analyzer, Model 121. Molar ratios were referred to Gly=1.00. Elemental analyses were performed by Galbraith Laboratories, Inc. Knoxville, Tenn. The analytical results for the elements indicated by their respective symbols were within $\pm 0.4\%$ of theoretical values. Optical rotations were measured with a Bellingham Stanley, Ltd., Model A polarimeter, type pl.

EXAMPLE 1

β -(S-Benzylmercapto)- β , β -cyclopentamethylenepropionyl-Tyr(Me)-Phe-Gln-Asn-Cys(Bzl)-Pro-Arg(Tos)-Gly-NH₂

(a) Combination of Solid Phase and Solution Methods
Boc-Tyr(Me)-Phe-Gln-Asn-Cys(Bzl)-Pro-Arg(Tos)-Gly-NH₂, prepared by the method of Bankowski et al., *J. Med. Chem.*, vol. 21 (1978) at 850 (319 mg, 0.26 mmol), was dissolved in TEA (6.5 ml) and stirred at room temperature for 40 mins. Cold ether (20 ml) was added to produce a precipitate which was filtered and washed with ether (5 \times 10 ml). The product was dried in vacuo over sodium hydroxide pellets. This material (318.5 mg) was dissolved in DMF (0.8 ml), to which was added N-methylmorpholine (10 μ l). The resulting solution had a pH of 7–8, measured with moist pH paper. After this neutralized solution was stirred at room temperature for 30 mins, a solution of p-nitrophenyl β -(S-benzylmercapto)- β , β -cyclopentamethylenepropionate, Nestor et al., *J. Med. Chem.*, vol. 18 (1975) at 284, (445 mg, 1.155 mmol in 0.4 ml of DMF) was added. The reaction mixture was stirred at room temperature. After 72 hours' stirring, TLC analysis using system D showed that the reaction mixture still contained a trace of the free octapeptide amide. N-Hydroxybenzotriazole monohydrate, König et al., *Chem. Ber.*, vol. 103 (1970) at 788, (39.3 mg, 0.26 mmol) was added. Coupling was complete within 5 hours. The precipitate was filtered, washed with cold ethyl acetate (4 \times 10 ml) and dried in vacuo. The crude product (339 mg) was twice reprecipitated from DMF-methanol to give the acylpeptide amide (295.2 mg, 77.3%): mp. 209°–211° C.; $[\alpha]_D^{24} = -43.6^\circ$ (C 0.5, DMF); $R_f(E)$ 0.45, $R_f(F)$ 0.63. Anal. (C₇₃H₉₄O₁₄N₁₄S₃) C, H, N.

Amino acid analysis: Tyr, 0.80; Phe, 1.01; Glu, 1.04; Asp, 1.02; Cys(Bzl), 0.98; Pro, 1.06; Arg, 1.01; Gly, 1.00; NH₃ 2.91.

(b) Total Synthesis on Resin

Boc-Tyr(Me)-Phe-Gln-Asn-Cys(Bzl)-Pro-Arg(Tos)-Gly-resin (1.11 g, 0.4 mmol prepared from Boc-Gly-resin using solid phase methodology) was converted to the acyloctapeptide resin (1.167 g, weight gain 57 mg, 97.6% of theory) in one cycle of deprotection, neutralization and coupling with p-nitrophenyl β -(S-benzylmercapto)- β , β -cyclopentamethylenepropionate, see Nestor supra. The resin was ammonolyzed, Manning, *J. Am. Chem. Soc.*, vol. 90 (1968) at 1348. The product was extracted with dimethylformamide (DMF). After the solvent was evaporated in vacuo, the residue was pre-

cipitated by addition of water. The crude product (410 mg) was twice reprecipitated from DMF-ethanol to give the acyloctapeptide (302 mg, 50.7% based upon initial glycine content of the resin); mp. 206°–208° C. (decomp); $R_f(E)$ 0.45, $R_f(F)$ 0.63; $[\alpha]_D^{24} = -43.1^\circ$ (C 1, DMF). Anal. (C₇₃H₉₄N₁₄O₁₄S₃) C, H, N.

Amino acid analysis: Tyr, 0.79; Phe, 1.01; Glu, 1.03; Asp, 1.04; Cys(Bzl), 0.97; Pro, 1.03; Arg, 0.99; Gly, 1.00; NH₃, 2.95.

EXAMPLE 2

β -(S-Benzylmercapto)- β , β -cyclopentamethylenepropionyl-Tyr(Bzl)-Phe-Gln-Asn-Cys(Bzl)-Pro-Arg(Tos)-Gly-NH₂

Boc-Tyr(Bzl)-Phe-Gln-Asn-Cys(Bzl)-Pro-Arg(Tos)-Gly-resin (1.46 g, 0.5 mmol) was converted to acyloctapeptide resin (1.55 g, weight gain 70 mg, 95.9% of theory) as in Example 1 by one cycle of deprotection, neutralization and coupling with p-nitrophenyl β -(S-benzylmercapto)- β , β -cyclopentamethylenepropionate. The product obtained by ammonolysis of the resin was extracted with DMF. The solvent was evaporated in vacuo and the residue was precipitated by addition of water. The crude product (723 mg) was reprecipitated from DMF-ethanol and DMF-2% aqueous AcOH. (488 mg; 62.4% based on initial Gly content on the resin); mp. 183°–185° C.; $R_f(E)$ 0.38; $R_f(D)$ 0.41; $[\alpha]_D^{23} = -32.9^\circ$ (C 1 DMF). Anal. (C₇₉H₉₈N₁₄O₁₄S₃) C, H, N.

Amino acid analysis: Tyr, 0.97; Phe, 1.02; Glu, 1.05; Asp, 1.01; Cys(Bzl), 0.98; Pro, 1.04; Arg, 0.98; Gly, 1.00; NH₃.

EXAMPLE 3

[1-(β -Mercapto- β , β -cyclopentamethylenepropionic acid), 2-(O-methyl)tyrosine]-arginine vasopressin

(a) From Nonapeptide Amide

A solution of the protected nonapeptide amide, prepared as in Example 1, (170 mg, 0.114 mmol) in 400 ml of ammonia (dried over sodium and redistilled) was stirred at the boiling point with sodium from a stick of the metal contained in a small bore glass tube until a light blue color persisted in the solution for 30 sec, in accordance with duVigneaud, *J. Am. Chem. Soc.*, vol 76 (1954) at 3115. Dry glacial acetic acid (0.4 ml) was added to discharge the color. The solution was evaporated. A solution of the residue in aqueous acetic acid (0.2%; 800 ml), was treated with 2 M ammonium hydroxide solution to give a solution of pH 7.5. To this stirred solution was added gradually an excess of a solution of potassium ferricyanide (0.01 M, 11.4 ml), Hope et al., *J. Biol. Chem.*, vol. 237 (1962) at 1563. The yellow solution was stirred for 90 min more and for 1 h with anion-exchange resin (BioRad AG-3, Cl⁻ form, 10 g damp weight). The suspension was filtered slowly through a bed of resin (80 g damp weight). The resin bed was washed with 300 ml of aqueous 0.2% acetic acid and the combined filtrate and washings were lyophilized. The resulting powder (1386 mg) was desalted on a Sephadex G-15 column (110×2.7 cm) and eluted with aqueous acetic acid (50%) at a flow rate of 4 ml/h by the technique of Manning et al., *J. Chromatog.*, vol. 38 (1968) at 396. The eluate was fractionated and monitored for absorbance of 280 nm. The fractions comprising the major peak were pooled and lyophilized. The residue (55.5 mg) was further subjected to gel filtration on a Sephadex G-15 column (100×1.5 cm) and eluted with aqueous acetic acid (0.2 M) at a flow

rate of 2.5 ml/h. The peptide was eluted in a single peak (absorbance 280 nm). Lyophilization of the pertinent fractions yielded the vasopressin analog (49 mg, 37.3%) $R_f(E)$ 0.19; $R_f(F)$ 0.30; $[\alpha]_D^{22} = -59.6$ (C 0.19, 1 M AcOH).

Amino acid analysis: Tyr, 0.81; Phe, 1.01; Glu, 1.04; Asp, 0.98; Pro, 1.04; Arg, 0.95; Gly, 1.00; NH₃, 3.10. Analysis following performic acid oxidation prior to hydrolysis according to Moore, *J. Biol. Chem.*, vol. 238 (1963) at 235, gave a Cys(O₃H)-Gly ratio of 1.03:1.00.

(b) From Acyloctapeptide

Treatment of the acyloctapeptide (160 mg, 0.107 mmol) as described in Example 3 (a) yielded the analog (64 mg, 51.7%), which was indistinguishable from the foregoing preparation by TLC: $[\alpha]_D^{23} = -59.1^\circ$ (C 0.5, 1 M AcOH).

Amino acid analysis: Tyr, 0.80; Phe, 1.02; Glu, 1.02; Asp, 0.98; Pro, 1.03; Arg, 0.96; Gly, 1.00; NH₃, 3.05. Analysis following performic acid oxidation prior to hydrolysis gave a Cys-(O₃H)-Gly ratio of 1.02:1.00.

EXAMPLE 4

[1-(β -Mercapto- β , β -cyclopentamethylenepropionic acid] arginine vasopressin

Treatment of the acyloctapeptide (173 mg, 0.111 mmol) as described in Example 3 (a) yielded the analog (66 mg, 52%) $R_f(E)$ 0.19, $R_f(F)$ 0.43; $[\alpha]_D^{23} = -58.7^\circ$ (C 0.5, 1 M AcOH).

Amino acid analysis: Tyr, 0.96; Phe, 0.98; Glu, 1.01; Asp, 1.01; Pro, 1.05; Gly, 1.00; NH₃, 2.95. Analysis following performic acid oxidation prior to hydrolysis gave a Cys(O₃H)-Gly ratio of 1.01:1.00.

EXAMPLE 5

[1-(β -Mercapto- β , β -cyclopentamethylenepropionic acid), 2-substituted, 4-valine]-(L- and D-)-arginine vasopressin

Compounds of this series were prepared by solid-phase synthesis, modified as in Manning et al., *J. Med. Chem.*, vol. 16 (1973) at 975, Kruszynski et al., *J. Med. Chem.*, vol. 23 (1980) at 364 Manning et al. *J. Med. Chem.*, vol. 24 (1981) at 701, to obtain protected intermediates for each analog. The procedures of Bodanszky et al., *J. Am. Chem. Soc.*, vol. 81 (1959) at 5688 and *J. Org. Chem.*, vol. 39 (1974) at 444, employing a p-nitrophenyl ester, facilitated by the use of hydroxybenzotriazole (Konig et al., supra), were used for the coupling of β -(S-benzylmercapto)- β , β -cyclopentamethylenepropionic acid in accordance with Nestor, supra, to obtain precursor compounds. Each precursor was deblocked (duVigneaud, supra) with sodium in liquid ammonia. The resulting disulfhydryl compounds were oxidatively cyclized with potassium ferricyanide (Hope et al., supra). The analogs were desalted and purified by gel filtration on Sephadex G-15 by a two step procedure using 50% acetic acid and 0.2 M acetic acid, respectively, as eluants. The purity and identity of each analog was ascertained by thin-layer chromatography in two different solvent systems, Kruszynski et al., *J. Med. Chem.*, vol. 23 (1980) at 364, or by amino acid analysis as above.

Compounds of Formula I, or related to Formula I, prepared by the foregoing procedure were assayed by TLC on silica gel in two solvent systems: E. butanol/a-

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6. [1-(β-Mercapto-β,β-cyclopentamethylenepropionic acid), 2-D-leucine, 4-valine]-arginine vasopressin, a compound of claim 1.

7. [1-(β-Mercapto-β,β-cyclopentamethylenepropionic acid), 2-D-isoleucine, 4-valine]-arginine vasopressin, a compound of claim 1.

8. [1-(β-Mercapto-β,β-cyclopentamethylenepropionic acid), 2-D-phenylalanine, 4-valine, 8-D-arginine] vasopressin, a compound of claim 1.

9. A method for antagonizing the in vivo response of an animal to the antidiuretic action of an antidiuretic hormone, comprising administering to the animal being treated an amount of a compound of claim 1, in admixture with a physiologically and pharmaceutically acceptable carrier, effective to antagonize the antidiuretic response to the antidiuretic hormone.

10. The method of claim 9, wherein the antidiuretic hormone is arginine vasopressin.

11. The method of claim 9, wherein the compound is administered parenterally.

12. A method for antagonizing the in vivo response of an animal to the antidiuretic action of an antidiuretic hormone, comprising administering to the animal being treated an amount of the compound of claim 7, in admixture with a physiologically and pharmaceutically acceptable carrier, effective to antagonize the antidiuretic response to the antidiuretic hormone.

13. A method for antagonizing the in vivo response of an animal to the antidiuretic action of an antidiuretic hormone, comprising administering to the animal being treated an amount of the compound of claim 4, in admixture with a physiologically and pharmaceutically acceptable carrier, effective to antagonize the antidiuretic response to the antidiuretic hormone.

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